RESTORATION OF ABANDONED MINE SITES

FINAL WORK PLAN

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July 2002

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LIST OF ACRONYMS & ABBREVIATIONS

AES Atomic Emission Spectroscopy
APP Acid Producing Potential
asap As Soon As Possible

ASTM American Society for Testing and Materials

bgs below ground surface

CaCO₃ Calcium Carbonate

CEC Cation Exchange Capacity

CENWD U.S. Army Corps of Engineers – Northwestern Division

CENWO U.S. Army Corps of Engineers – Omaha District

CEPOD U.S. Army Corps of Engineers – Pacific Ocean Division
CERCLA Comprehensive Environmental Response Compensation and

Liability Act

CESPD U.S. Army Corps of Engineers – South Pacific Division
CESPA U.S. Army Corps of Engineers – Albuquerque District
CESPK U.S. Army Corps of Engineers – Sacramento District

CH Fat Clay
CL Lean Clay
cm Centimeter
CN Cyanide

COC Chain of Custody

CSM Colorado School of Mines CVAA Cold Vapor Atomic Absorption

DI Deionized

DMG Division of Minerals and Geology

DO Dissolved Oxygen

DQCR Daily Quality Control Report
DQO Data Quality Objective
DRI Desert Research Institute

ECB Environmental Chemistry Branch
EE/CA Engineering Evaluation/Cost Analysis

Eh Redox or Oxidation Potential

EM Electromagnetic

GC/MS Gas Chromatography/Mass Spectroscopy

GPR Ground-Penetrating Radar
GPS Global Positioning System

GSSI Geophysical Survey Systems, Inc.

 $\begin{array}{ccc} HCN & Hydrogen Cyanide \\ H_2O_2 & Hydrogen Peroxide \end{array}$

ICP Inductively Coupled Plasma

ID Identification
I.D. Inner Diameter

IDW Investigative-Derived Waste

IH Industrial Hygienist

"J" Estimate Code

K Hydraulic Conductivity

LCS Laboratory Control Sample

LCSD Laboratory Control Sample Duplicate

LIMS Laboratory Information Management System LQMM Laboratory Quality Management Manual

LRL Laboratory Reporting Limit

MDL Method Detection Limit

ML Silt
mL Milliliter
mm Millimeter
MSL Mean Sea Level

MS/MSD Matrix Spike/Matrix Spike Duplicate

MW Monitoring Well

MWMP Meteoric Water Mobility Procedure

NaOH Sodium Hydroxide NAD North American Datum

NGVD National Geodetic Vertical Datum

NIST National Institute for Standards and Technology

NTU Nephelometric Turbidity Unit

O.D. Outside Diameter

OSHA Occupational Safety and Health Act
ORP Oxidation Reduction Potential

pH A measure of the Hydrogen Ion Concentration of materials

PM Project Manager ppm Parts Per Million PQL Practical Quantitation Limit psi Pounds per square inch PVC Polyvinyl Chloride

QA Quality Assurance

QAPP Quality Assurance Project Plan

QC Quality Control

QMP Quality Management Plan

"R" Unusable Code R Percent Recovery

RAMS Restoration of Abandoned Mine Sites

RL Reporting Limit

RPD Relative Percent Difference

SB Soil Boring

SB/MW Soil Boring/Monitoring Well

SD Sediment

SIR-2 Subsurface Interface Radar System-2 SOP Standard Operating Procedure

SPLP Synthetic Precipitation Leaching Procedure

SPT Standard Penetration Test

SS Surface Sample

SSA Site Specific Addendum
SSHO Site Safety and Health Officer
SSHP Site Safety and Health Plan

SW-# Solid Waste SW Surface Water

TB Trip Blank

TCLP Toxicity Characteristic Leaching Procedure

TDS Total Dissolved Solids TOC Total Organic Carbon

USACE United States Army Corps of Engineers
USCS Unified Soil Classification System

USEPA United States Environmental Protection Agency

VOC Volatile Organic Compounds

WAD Weak Acid Dissociable

WB Water Blank

WES Waterways Experiment Station

WHC Water Holding Capacity
WRDA Water Resources Development Act

1. INTRODUCTION

USACE has been provided authority for Restoration of Abandoned Mine Sites (RAMS) through the Water Resource Development Act (WRDA) 1999 section 560. This program is a regionally focused and stakeholder responsive program for the restoration of abandoned and inactive non-coal mines where water resources (ecosystem/habitat) have been degraded by past mining practices. This authority is intended to allow the Corps of Engineers to provide support to agencies that manage lands impacted by past mining. The scope of work for the Corps under RAMS will be developed in consultation with federal, state, and local government agencies and stakeholders to ensure that the stakeholder needs are being met and avoid duplication of effort and expenditures.

This Work Plan describes the field investigation activities, procedures, organization, objectives, functional activities and specific Quality Control (QC) activities to be used by the U.S. Army Corps of Engineers (USACE) RAMS program for Omaha District (CENWO), USACE Albuquerque District (CESPA), USACE Sacramento District (CESPK), and potentially other USACE districts within the boundaries of Northwestern Division (CENWD), South Pacific Division (CESPD), and Pacific Ocean Division (CEPOD). This Work Plan will provide the requirements to be followed in assessing the physical and environmental impacts from non-coal mining wastes.

Sites being investigated under the RAMS Program are not initially subject to the requirements of the Comprehensive Environmental Response Compensation and Liability Act (CERCLA), but may eventually come under the jurisdiction of the U.S. Environmental Protection Agency (USEPA) or Federal land management agencies. Therefore, work plans prepared for RAMS projects are of sufficient quality for use within the CERCLA process. The RAMS Work Plan is intended to provide standard information that will be applicable to most RAMS investigations. A Site-Specific Addendum (SSA) to this Work Plan will be prepared for each specific project. The SSA will reference the RAMS Work Plan and Site Safety and Health Plan (SSHP) and will include any site-specific requirements not included in the Work Plans.

1.1 PROJECT DESCRIPTION

The work described in the appendices includes the collection of surface soil and waste rock samples, advancement of soil borings for the collection of subsurface soil samples, the collection of surface water/sediment samples, the installation of monitoring wells for the collection of ground-water samples, performance of geophysical surveys, and performance of ground-water well pumping tests and slug tests to evaluate the vertical and lateral extent of potential contamination.

There are four appendices to this Work Plan:

- Appendix A Standard Operating Procedures (SOPs)
- Appendix B Field Equipment Operation Manuals
- Appendix C American Society for Testing and Materials (ASTM) Methods
- Appendix D Chemistry Quality Assurance Branch Laboratory Quality Management Manual (LQMM)

Field work will be conducted in accordance with the following Standard Operating Procedures (SOPs) (See Appendix A), if applicable.

- A1 Surface Soil/Rock Sampling Equipment and Procedures
- A2 Drilling Equipment and Procedures
- A3 Subsurface Soil/Rock Sampling Equipment and Procedures
- A4 Soil/Rock Homogenization Equipment and Procedures
- A5 Lithologic Description of Surface and Subsurface Soil Samples
- A6 Boring Log Completion
- A7 Investigative Derived Waste Procedures
- A8 Monitoring Well Design, Installation, and Abandonment Procedures
- A9 Monitoring Well Development Equipment and Procedures
- A10 Ground Water Sampling Equipment and Procedures
- A11 Surface Water and Sediment Sampling Equipment and Procedures
- A12 Equipment Decontamination Procedures
- A13 Sample Handling, Documentation, and Tracking Procedures
- A14 Field Documentation
- A15 Geophysical Survey Equipment and Procedures
- A16 Geotechnical Sampling Equipment and Procedures
- A17 Percolation Test Equipment and Procedures
- A18 Pumping Test Equipment and Procedures
- A19 Slug Test Equipment and Procedures

These SOPs will be referenced by number as they apply to the areas discussed in the SSAs.

Additional information is included in the SSHP regarding procedures required to ensure the health and safety of personnel performing field investigation duties. These documents also include discussions of activities to be performed and the interpretation and use of the data obtained during investigations.

The site-specific projects may involve the following tasks:

- Preparation of Site Specific Addenda to this Work Plan,
- Preparation of Site Specific Addenda to the SSHP,
- Collection of surface soil environmental samples,
- Collection of waste rock samples,
- Collection of surface water/sediment environmental samples
- Completion of soil borings for the collection of surface and subsurface soil environmental samples,
- Installation and development of new monitoring wells,
- Collection of environmental samples from new and/or existing monitoring wells,
- Off-site laboratory analysis of environmental samples to quantitatively assess contamination,
- Collection of geotechnical samples in support of designs for mitigation,
- Performance of a percolation test,

- Performance of geophysical surveys,
- Performance of ground water well pumping tests and slug tests to quantify hydrologic parameters,
- Performance of topographical surveys of affected areas,
- Design of grading and drainage plans,
- Preparation of Engineering Evaluation/Cost Analysis (EE/CA) reports,
- Collection of data to populate a RAMS database,
- Evaluation from a "watershed" perspective,
- Pursuit and promotion of innovative technologies for mine waste characterization and remediation,
- Support for the efforts of other federal agencies,
- Soil geotechnical testing,
- Percolation and/or infiltration testing, and
- Preparation of Reports.

1.2 OVERALL PROJECT OBJECTIVES

Specific project objectives will be jointly determined by project partners, stakeholders, and the USACE virtual team for each project and stated in the Site Specific Addendum. The general objective of the RAMS program is to support federal, state, and local stakeholders in their objectives for investigation and/or restoration of abandoned mines and to avoid duplication of effort by all participants.

1.3 PROJECT SCHEDULE

A project schedule will be provided in the SSA for each project.

2. SAMPLING LOCATIONS AND REQUIREMENTS

2.1 SAMPLING LOCATIONS

2.1.1 Surface Soil/Waste Rock Sampling Locations

The purpose of the surface soil/waste rock sampling is to determine contaminant concentrations the lateral extent of contamination in surface soils. (Surface soil sampling at soil boring locations is discussed under Soil Boring/Monitoring Well Sampling Locations in Section 3.1.2 below.) The selection of surface soil sampling locations will be based on the following criteria:

- locations within potential areas of contamination (lateral extent),
- locations down gradient topographically from potential areas of contamination (direction of migration), and
- locations in a topographic low area (following minor transportation by wind or water and/or evaporation).

2.1.2 Soil Boring/Monitoring Well Locations

The purpose of the soil borings/monitoring wells is to determine the vertical and lateral extent of surface and subsurface soil and/or ground-water contamination. The selection of soil boring/monitoring well locations will be based on the following criteria:

- locations within areas of potential contamination (vertical and lateral extent),
- locations down-gradient topographically from potential areas of contamination (direction of migration),
- locations up-gradient topographically from potential areas of contamination identified during previous investigations (up-gradient vertical and lateral extent), and
- locations within surface drainage ways or topographic lows (greater potential for ground water and area of concentration following minor transportation and/or evaporation).

2.1.3 Surface Water/Sediment Sampling Locations

The purpose of the surface water/sediment sampling is to determine the presence or absence of contamination related to mining activities. The selection of surface water sampling locations will be based on the following criteria:

- locations within surface drainage ways (potential presence of sediment and water),
- locations up-gradient topographically from potential areas of contamination identified during previous investigations (up-gradient vertical and lateral extent),
- locations in topographic low (greater potential for surface water during the dry, summer season), and

• locations down-gradient topographically from the source areas within the site (direction of migration).

2.2 SAMPLING REQUIREMENTS

2.2.1 Surface and Subsurface Soil Sampling Requirements for Soil Borings

Surface soil samples will be collected from the intervals indicated in the SSA. Soil analytical parameters are site-specific and are identified in the SSA.

2.2.2 **Ground Water Sampling Requirements**

Ground water analytical parameters are site-specific and are identified in the SSA. Lists of the potential analytes are presented in section 8 of this document.

2.2.3 Surface Water and/or Sediment Sampling Requirements

To prevent introduction of sampler-induced turbidity or sampler-caused sedimentation, surface water and sediments will be collected from downstream to upstream locations and the surface water sample will be collected prior to the collection of sediment samples.

3. FIELD QUALITY CONTROL

The following sections discuss the Field Quality Control Procedures that will be utilized for these projects.

3.1 FIELD ANALYTICAL PROCEDURES

The procedures for field measurement of pH, conductivity, turbidity, water level, and temperature are described in the appropriate SOPs in Appendix A of this Work Plan.

3.2 FIELD QUALITY CONTROL CHECKS

During field activities, USACE personnel will collect soil, ground water, surface water, and sediment duplicate samples for laboratory analysis. The SSA will specify the number of duplicate samples to be collected for each project.

3.3 FIELD DATA REDUCTION AND VALIDATION PROCEDURES

All field measurements (e.g. pH, conductivity, and temperature) will be directly read from the equipment, and field data reduction procedures will be minimal. Calibration will be performed in accordance with the manufacturer's recommendation. The calibration data and sample measurement data will be recorded in field log books or on data sheets as described in the appropriate SOPs immediately after measurements are taken. If recording errors are made, the error will be crossed out using a single line, initialed, and dated by the field member, and the correct information will be written in the space adjacent to the original (erroneous) entry.

After completing the sampling, the field data package (field logs, chain-of-custody forms, etc.) will be reviewed by the project team for completeness and accuracy. Items included in the validation of field data are listed below:

- Review of completeness of field data recorded on sampling logs,
- Verification that field duplicates were properly prepared, identified, and collected at the required frequency,
- Review of field analyses, equipment calibration, and equipment condition,
- Review of chain-of-custody (COC) forms for proper completion, dates, and signatures of field personnel,
- Review of the boring logs for completeness and consistency with COC paperwork, and
- Review of all other logs and/or forms for completeness and consistency with COC paperwork.

3.4 FIELD DATA REPORTING

Field data reporting will be conducted principally through the transmission of boring logs, monitoring well construction diagrams, well development logs, water sampling logs, field log books, data sheets,

and Daily Quality Control Reports (DQCR). Documentation of field calibration activities will be included in the field log books or on data sheets as described in the appropriate SOP. (Refer to SOP A14 of Appendix A).

3.5 FIELD PERFORMANCE AUDITS

If field performance audits are required in the SSA, they will meet the following requirements. A Quality Management Plan (QMP) will be prepared for each specific project and will indicate which performance audits will be performed and which project team member(s) will have those responsibilities. Audits are planned and documented evaluations of project operations that determine the adequacy and effectiveness of, as well as compliance with, the project plans. Performance and systems audits of both field and laboratory activities may be conducted to verify that field sampling and analyses, as well as off-site chemical analyses, are performed in accordance with the Work Plan.

Performance audits verify the laboratory's ability to correctly identify and quantify contaminants of concern in blind samples and as such, assess the accuracy of the measurement system. Laboratory performance audits will be performed by and at the discretion of USACE.

The objectives of performance audits are to:

- ensure all Work Plan and SSA requirements are being implemented,
- assess the effectiveness of the Work Plan and SSA
- verify that corrective actions are made in response to identified deviations

Systems audits evaluate the entire process of generating the environmental measurement from sample collection through entry of the analytical value into the project database. Systems audits are initially performed shortly after systems are operational and then on a schedule during the project lifetime.

The objectives of systems audits are to:

- ensure Work Plan and SSA are in use and understood.
- verify that required documentation is accurate and complete,
- assess the effectiveness of the Work Plans and identify necessary deviations, and
- verify the effectiveness of corrective actions, if required.

Field system audits will be conducted by the project team member(s) identified in the QMP who will be responsible for ensuring that corrective actions are complete and effective. The field team leader will be responsible for correcting all deficiencies cited as deviations from the Work Plans.

An initial field systems audit will be conducted at a frequency identified in the QMP.

The primary purpose of the field systems audit is to verify that the field team is complying with the DQOs and properly documenting field activities. Examination of field sampling records, field instrument

calibration, proper log book or data sheet documentation, in-field custody procedures, sample container storage and labeling, packaging and shipping procedures, and observation of sample collection and handling will be included. All field systems audits will consist of the following:

- Pre-field mobilization preparation to familiarize the field team with the specific forms, field equipment
 calibration protocols, logs, and instrument records to be kept during field activities, and their
 importance,
- Inspection of field records with on-site identification of documentation gaps, inconsistencies, and deviations from the Work Plan,
- Observation of field sampling with immediate notification to the Project Geologist of deviations from the Work Plan,
- Discussions of the audit findings with the geologist at the site and notification of the audit findings to the USACE Project Manager, and
- Post-audit activities including preparation of an audit report detailing deviations, non-conformances, and corrective action recommendation.

3.6 SPECIFIC PROCEDURES

If identified in the QMP for each specific project, the following procedures will be used to assess field data precision, accuracy, and completeness.

3.6.1 Field Measurements

Accuracy of the field measurements will be assessed using daily instrument calibration and calibration checks.

3.6.2 Field Corrective Actions

If it is determined that a situation will result in nonconformance with the project plans and will require corrective action, a nonconformance report will be initiated by the project team. For RAMS projects, the project team will include the project manager, project technical staff, and project field team leader. The nonconformance report will document what the nonconformance consists of, what corrective action will be taken, and who is responsible for implementing and reporting the implementation of the corrective action.

The project team will be responsible for corrective actions that include:

- Evaluating all reported non-conformances,
- Controlling additional work on nonconforming items,
- Determining the corrective action to be taken,
- Directing the field staff to implement the corrective action,
- Maintaining a log of non-conformances,
- Reviewing nonconformance reports and corrective actions taken, and

• Ensuring nonconformance reports are included in the project files.

The project team will ensure that no additional work that is dependent on the nonconforming activity is performed until corrective actions are completed.

Corrective actions for field measurements may include:

- Repeating the measurement,
- Checking that proper adjustments have been made for ambient conditions such as temperature,
- Checking the batteries of the sampling equipment,
- Checking the calibration of the sampling equipment,
- Replacing the instrument or measurement devices, or
- Stopping the work.

The field team leader is responsible for all on-site activities, unless otherwise indicated in the SSA. The field team leader is required to implement corrective actions. The change in the program will be documented in the project files. The field team leader must agree with the change prior to field implementation. Actions taken prior to the change will be evaluated to determine the significance of the data obtained and its effect on the data quality objectives for the project.

The project team is responsible for controlling, tracking, and ensuring the implementation of the changes. Reports on all changes will be distributed to all affected parties. The USACE Project Manager will be notified within 24 hours that a field corrective action has occurred and the action taken.

3.7 OFF-SITE ANALYSES

Laboratories that will perform chemical analytical analyses associated with these projects include, but are not limited to:

U.S. Army Corps of Engineers
Chemical and Material Quality Assurance Branch Laboratory
420 South 18th Street
Omaha NE 68102
Telephone (402) 444, 4314

Telephone: (402) 444-4314 Point of Contact: Laura Percifield

Energy Laboratories 1120 So. 27th Street Billings MT 59101

Telephone: (800) 735-4489 Point of Contact: John Standish

Colorado School of Mines (CSM)

(to be supplied in Site Specific Addendum if this laboratory will be utilized for the project)

Additional qualified analytical laboratories may be identified for use at specific sites. If a laboratory not identified in this plan will be used, information regarding the laboratory will be included in the SSA.

The following geotechnical laboratory will be involved with this project: (to be supplied in Site Specific Addendum if geotechnical samples will be collected for the project)

An Environmental Chemistry Branch (ECB) Laboratory Information Management System (LIMS) number will be assigned to each sample submitted for analysis. A separate LIMS number will be assigned to each project and will be included in the SSA. The LIMS number must be added to the labels and COC records for all samples shipped to the ECB Laboratory for projects being performed under this Work Plan.

4. CHEMICAL DATA QUALITY OBJECTIVES

4.1 DATA USES

DQOs are qualitative and quantitative statements that specify the objectives of the project, define the types of decisions that will be made, identify the intended use of the data and design a data collection program which addresses the quantity and quality of data required to support the project objectives. The DQOs are based on the end uses of the data to be collected and may include:

- determining if detected compounds are site-related contamination rather than representative of naturally-occurring concentrations,
- determining the vertical and lateral extent of the contamination in surface soils, subsurface soils, surface water/sediments, and ground water,
- refining conceptual models that may be used to determine exposure pathways for biological/ecological receptors for each site,
- collecting data of sufficient quality to support a quantitative assessment of carcinogenic and non-carcinogenic risks to human receptors as well as support a quantitative assessment of risk to ecological receptors, if necessary,
- collecting data of sufficient quality to support fate and transport modeling of contamination, if necessary,
- collecting data of sufficient quality to support a feasibility study and selection of a remedial option or options, if necessary, and
- determining the need and scope for a pre-design investigation, if necessary.

4.2 DATA QUALITY NEEDS

Sites being investigated under the RAMS program usually are not initially subject to the requirements of CERCLA but may eventually become subject to CERCLA and under the jurisdiction of the USEPA. Therefore, the work plans being prepared for RAMS are intended to ensure that data of sufficient quality for use within the CERCLA process is being obtained during the RAMS investigations. All QA/QC procedures described herein are generally structured in accordance with applicable technical standards and the USEPA requirements, regulations and guidance.(duplication is very important for this info!)

Definitive data as defined by the EPA uses rigorous analytical methods with satisfactory QA/QC requirements such as approved EPA reference methods are required for these projects. Data will be analyte-specific, with confirmation of analyte identity and concentration. Proper methods provide the necessary guidelines to produce tangible raw data (e.g., chromatograms, spectra, digital values) in the form of paper printouts or computer-generated electronic files. Data may be generated at the site or at an off-site location, as long as the QA/QC requirements are satisfied. For these projects, data will generally be generated at an off-site laboratory.

Screening data are generated by rapid, less precise methods of analysis with less rigorous sample preparation. Screening data are often used for health and safety monitoring or initial site characterization to locate areas for subsequent analyses.

4.3 METHOD SPECIFIC DATA QUALITY OBJECTIVES

Precision, accuracy, representativeness, completeness and comparability goals for this project are described below. These data quality parameters are expressed as goals due to uncertainties regarding field conditions and the complexity of the sample matrices at the site. The ability of the laboratory to extract and quantify the target analytes is dependent on the matrix complexity. The variability of field conditions, such as soil type and contaminant deposition, also will affect the data representativeness and comparability.

4.3.1 Precision

Laboratory: Precision is a measure of the degree of mutual agreement among data that have been collected the same way under prescribed conditions. Precision is assessed by replicate measurements of known standards and analysis of duplicate field samples. Precision is expressed as relative percent difference (RPD) between duplicate samples. Site-specific matrix spike/matrix spike duplicate (MS/MSD) pairs will be used to assess analytical precision, and field duplicates will be used to assess both analytical and sampling precision. Data with detected concentrations greater than three times the method detection limit (MDL) will be used to assess precision. The number of MS/MSDs collected and analyzed will generally be 5 to 10% of the total number of samples. Field precision will be determined by the RPD of analyte in the original and duplicate samples:

Field: Field sampling precision will be assessed through comparisons of field duplicate sample analytical results. The number of field duplicate samples collected will generally be equivalent to 10% of the number of original samples collected.

Precision for field measurements will be determined by performing replicate measurements on the same sample (i.e., a second measurement will be performed on 10% of the field samples collected). Comparison of consecutive measurements will be used to help determine the precision of the field sampling. When measurements are within 10% of each other, precision of field measurements will be considered adequate.

4.3.2 Accuracy

Laboratory: Accuracy is the measure of the proximity of an observed value to an accepted value. Accuracy is expressed as percent recovery for each analyte of the spike samples (MS/MSDs, LCS/LCSDs, surrogates, etc.).

R = [(Spike sample result - sample result)/Spike concentration added] x 100

where: %R = percent recovery

Field: Field accuracy is defined as the collection of a sample that truly reflects the quality of the media sampled, is adequately preserved and is not biased by field-induced contamination during sample collection, handling and shipment. Field accuracy will be assessed by analyzing trip blanks, laboratory checks of pH for preserved water samples, and the temperature of the samples when received by the laboratory. Samples will be maintained at a temperature of 4° C +/- 2° C.

Field Blanks

Field blank samples are not required with soil or sediment sampling.

New tubing and/or disposable bailers will be used for each ground-water well sampling event. This will reduce the probability of cross-contamination between wells and field blank samples will not be required with well water samples.

Trip Blanks

Trip blanks will accompany all sample shipments that contain water samples for Volatile Organic Compound (VOC) analysis. Trip blanks provide an indication of contamination attributable to sample collection, handling and shipment. Trip blanks will be prepared in the field from "carboys" of organic-free water provided by the USACE Environmental Chemistry Branch (ECB) Laboratory of Waterways Experiment Station. These blanks will be analyzed for all (VOC) analytes of concern. Trip blanks will not be required for soil samples.

Trip blanks will not contain detectable project analytes above the analytical laboratory reporting limits) (LRLs), and will have a proper pH and temperature when received by the laboratory. The sample LRL is the lowest concentration of an analyte that can be reported by the laboratory to be present in a sample with a specified level of confidence. Common contaminants arising from field sampling equipment, decontamination solutions and laboratory operations may be detected (these may include acetone, methylene chloride, toluene, 2-butanone, and/or common phthalates.) Trip blank samples will not contain these contaminant at concentrations greater than five times the LRL. Samples containing concentrations above these values will require the initiation of corrective action.

4.3.3 Representativeness

Laboratory: Representativeness expresses the degree to which data accurately and precisely represents the characteristics of a population of samples, parameter variations at a sampling point, a process or an environmental condition. Representativeness is a measure of how closely the measured results reflect the actual concentration or distribution of the chemical compounds in the sample. Representativeness of the analytical results can be assessed by comparing the results for field duplicate samples collected from a site. Some samples may require analysis of multiple phases to obtain representative results. To ensure representativeness in the laboratory, USACE will request that samples be prepared and analyzed together as much as possible so that the laboratory QC is applicable to this project's samples and laboratory variables are minimized.

Field: Under this Work Plan, field representativeness will be satisfied by ensuring that the Work Plan is followed, proper sampling techniques are used, proper sample handling and documentation procedures are followed, and proper analytical procedures are followed in the field. Proper documentation will establish that protocols have been followed and sample identification and integrity ensured.

4.3.4 Completeness

Laboratory: Completeness is defined as the amount of valid data obtained from the laboratory compared with the amount of data that would be expected under normal conditions. Completeness is presented in Table 4-1 as the percent of data that is valid. Data are considered valid if they are unqualified or if they are qualified as estimated. For analyses, the completeness of laboratory analytical data will be 90 percent or better. The goal for holding times is 100 percent.

The percent completeness for each set of samples can be calculated as follows:

Completeness = [valid data obtained/total data planned] $\times 100$

where valid data are determined by the data acceptance criteria defined in this Work Plan.

Field: Completeness of field activities will be measured as the amount of field measurements (pH, conductivity, temperature, headspace analysis, etc.) that comply with SOPs for calibration and precision requirements compared to the whole population of field measurements attempted. The field data completeness objective for this project is a minimum of 95% of all samples.

Field sample collection completeness will be measured as the total number of field samples of sufficient volume collected for laboratory analysis compared to the total number of samples planned for collection in the SSA. Field sample collection completeness for this project is expected to exceed 95% for soil samples. Ground water may not be present at all proposed monitoring well locations, and this may have a significant effect on the completeness of the ground-water sampling.

4.3.5 Comparability

Laboratory: Comparability is an expression of the confidence with which one data set can be compared with another. Comparability of analytical data will be achieved through the use of accepted and documented analytical methods. The data will be reported in units consistent with Federal and State regulations, methods, and guidelines. LRLs for all analytes will be recorded on all lab data report sheets for all samples within the sample set. Comparability between databases will be achieved by using standardized sampling and analysis methods and data formats.

Field: Comparability of data obtained during this investigation is dependent upon the proper design of the sampling program and will be satisfied by ensuring the Work Plan is followed and proper sampling and handling techniques are used. The intent is that field data will be directly comparable to data obtained during subsequent phases of the project.

4.3.6 Sensitivity

Sensitivity broadly describes the method detection/quantitation/LRLs established to meet the project-specific DQOs. The sample LRL is the lowest concentration of an analyte that can be reported by the laboratory to be present in a sample with a specified level of confidence The LRLs are a function of the sample characteristics, method quantitation, and laboratory performance.

Method quantitation limits are determined by the laboratory and defined as the level at which the laboratory can reliably quantitate the concentration of an analyte on multiple analyses. The LRLs can be equal to the method quantitation limits, but often are above since method quantitation limit studies are performed using laboratory-prepared samples (spiked deionized [DI] water); whereas, environmental samples are naturally more variable. LRLs and/or Practical Quantitation Limits (PQLs) are provided in Section 6.0. For this project, data reported below the LRLs will be reported as estimated. Factors that may result in elevated LRLs are discussed below.

- High concentrations of target or non-target analytes may require that the sample extract be
 diluted to avoid saturation of the detector or to quantify the analyte concentration within the
 calibration range of the instrument. Consequently, LRLs are elevated in proportion to the
 dilution factor.
- Matrix interferences may require that the sample be diluted to reduce or eliminate the interference. Consequently, the LRLs are elevated in proportion to the dilution factor.
- Soil analytical results and LRLs are corrected for the moisture content of the sample. The LRLs listed Section 5.0 for soils are based on wet weight. LRLs calculated by the

laboratory for soil on a dry weight basis will be higher. Because moisture content can vary from sample to sample, LRLs will vary accordingly.

- The physical characteristics of the matrix do not permit concentration to the required final volume during sample preparation resulting in a larger sample extract volume and consequently an elevation in LRLs.
- Matrix interference may require the LRLs be elevated because of the inability to quantify data below the elevated LRL

TABLE -1 COMPLETENESS GOALS

| Analyte | Matrix | Completeness |
|-------------------------------|--------|--------------|
| | | (%) |
| Holding Time for all analytes | Soil | 90 |
| Holding Time for all analytes | Water | 90 |
| VOCs | Soil | 90 |
| VOCs | Water | 90 |
| Metals | Soil | 90 |
| Metals | Water | 90 |
| Cyanide | Soil | 90 |
| Cyanide | Water | 90 |
| Water Quality Parameters | Water | 90 |

 $\label{eq:water_equality} Metals = Selected metals from Inductively Coupled Plasma - Atomic Emission Spectroscopy (ICP-AES) list \\ Water Quality Parameters/ = Alkalinity, Hardness (as CaCO_3), Conductivity, Total Dissolved Solids (TDS), Nitrate/Nitrite, Sulfate, Carbonate, Bicarbonate, Chloride and Ammonia \\ National Carbonate, Carbonate, Chloride and Ammonia \\ National Carbonate, Carbonate, Carbonate, Chloride and Carbonate, Carbonat$

5. PROJECT ORGANIZATION AND QC RESPONSIBILITY

5.1 OVERVIEW

The project management organization is designed to provide a line of responsibility and authority, supported by a management control structure. Basically, this control structure provides for:

- Clearly identified lines of communication and coordination
- Project budget and schedule monitoring
- Key technical resources management
- Periodic financial management and progress reports
- QC

5.2 RESPONSIBILITIES

USACE has responsibility for coordinating all phases of the fieldwork performed by USACE with project stakeholders. Some projects may involve other agencies, analytical facilities, and other field personnel. Where appropriate, USACE will provide project management, perform the field investigations, review and evaluate the data, and prepare draft and final reports. The project team will review all environmental data. All personnel involved in this project will adhere to COC and field documentation protocols and will perform all activities in accordance with the Work Plan and SSHP.

The various QA, field, laboratory and management responsibilities of key project personnel are defined below. Information regarding analysis of samples by the Environmental Chemistry Branch (ECB) Laboratory is provided in the ECB LQMM. A copy of this document is included as Appendix D.

5.3 MANAGEMENT RESPONSIBILITIES

The USACE Project Manager (PM) has overall funding responsibility for all phases of the investigation. The PM is responsible for coordinating the project QC reviews, scheduling, and data interpretation and has responsibility for implementing all phases of the investigation. In this capacity, the PM will coordinate with all RAMS project personnel during sampling activities and analyses of samples by ECB Laboratory and other laboratories for this work.

The PM's primary function is to ensure that technical, financial, and scheduling objectives are achieved. The PM will be the major point of contact internally and externally and control for all matters concerning the project.

5.4 QUALITY ASSURANCE RESPONSIBILITIES

The project team members will coordinate/implement the fieldwork to ensure that data collected will support the intent of the sampling effort, provide direction to the project field team and complete progress or other required management reports.

5.4.1 Project Chemist

The Project Chemist has primary responsibility for:

- for writing the appropriate sections of the Work Plan and SSA,
- ongoing surveillance of project activities to help ensure conformance with this Work Plan,
- for appropriate procedures, including strict QC and decisions about data acquisition,
- data quality issues,
- coordination of analytical requirements with laboratory analyses,
- initial data review of all sample results from the analytical laboratories,
- data review and evaluation as indicated in the SSA,
- and the preparation of the report..

5.4.2 Project Geologist and/or Project Engineer

The Project Geologist and/or Project Engineer has primary responsibility for:

- writing the appropriate sections of the Work Plan and SSA,
- ongoing surveillance of project activities to help ensure conformance with this Work Plan, and
- oversight of the fieldwork as indicated in the SSA.

5.4.3 Project Industrial Hygienist

The Project Industrial Hygienist (IH), in consultation with the rest of the project team, writes the SSHP.

The Project IH has primary responsibility for:

- writing or reviewing any necessary modification to the SSHP,
- providing technical assistance to the field team, and ensure
- ensuring that that the SSHP meets OSHA requirements.

The Project IH will also assist with reviewing analytical lab results for risk analysis purposes, if required by the SSA.

5.5 LABORATORY RESPONSIBILITIES

Samples for all analyses will be sent to the following laboratories:

- ECB Laboratory of the WES
- CSM
- Energy Laboratories
- Other qualified laboratories identified in SSA

In general, field QC samples, such as field duplicates, will be obtained at a frequency of 10% (i.e., on average, a QC sample will be obtained for every ten samples collected), unless otherwise indicated in the SSA. The laboratory will run lab QC samples (i.e., MS/MSD samples, etc.) at the required frequency of 5%, unless otherwise indicated in the SSA.

The laboratory will designate a Project or Program Manager, Operations Manager, QA Officer and Sample Custodian for the project. The functions of these individuals are explained in the respective laboratory QA manuals and are briefly described below. In some cases, titles other than those shown have been used by the laboratories.

5.5.1 <u>Laboratory Program Manager</u>

The Laboratory Program Manager will report to the Laboratory Director and will be responsible for:

- Ensuring all resources of the laboratory are available when required; and,
- Providing an overview of final analytical reports.

5.5.2 Laboratory Operations Manager

The Laboratory Operations Manager will report to the Laboratory Program Manager and will be responsible for:

- Coordinating laboratory analyses;
- Supervising in-house COC;
- Scheduling sample analyses;
- Overseeing data review;
- Overseeing preparation of analytical reports; and,
- Approving final analytical reports prior to submission to the USACE.

5.5.3 Laboratory Quality Assurance Officer

The Laboratory's QA Officer has overall responsibility for data after it leaves the bench. The Laboratory's QA Officer will be independent of the operating departments and will communicate data issues through the Laboratory's Program Manager to the Project Chemist. In addition, the Laboratory's QA Officer will:

- Overview laboratory QA,
- Overview QA/QC documentation,
- Conduct detailed data review.
- Determine whether to implement laboratory corrective actions if required,
- Notify the Project Chemist of data quality issues,
- Define appropriate laboratory QA procedures, and
- Prepare laboratory SOPs.

5.5.4 Laboratory Sample Custodian

Each Laboratory's Sample Custodian will report to the Laboratory's Operations Manager. Responsibilities of the Sample Custodian will include:

- Receiving and inspecting the incoming sample containers,
- Recording the condition of the incoming sample containers,
- Signing appropriate documents,
- Verifying COC reports and their accuracy,
- Notifying appropriate analytical staff of sample receipt and sample integrity issues (e.g., holding times),
- Assigning a unique identification number and customer number and entering each into the sample receiving log,
- With the help of laboratory staff, initiating and documenting transfer of the samples to appropriate lab sections, and
- Controlling and monitoring access/storage of samples and extracts.

Independent QA will be provided by the Laboratory's Program Manager and QA Officers prior to release of project data.

5.5.5 Laboratory Technical Staff

The Laboratories' Technical Staff will be responsible for continuance of COC, sample analysis, analytical documentation and identification of corrective actions, if necessary. Laboratory Technical Staff will report directly to the Laboratory Operations Manager or other appropriate Manager.

5.6 FIELD RESPONSIBILITIES

5.6.1 Field Team Leader

The Field Team Leader is responsible for the day-to-day field data collection and sampling activities of the various resource specialists under his/her supervision. The Field Team Leader is an experienced environmental professional (geologist, geological engineer, chemist, or other technical professional as identified in the SSA). Specific Field Team Leader responsibilities include:

- Day-to-day coordination of technical issues in specific areas of expertise
- Implementation field-related portions of the Work Plan
- Coordination and management of field staff during drilling, well installation and sampling
- Implementation of QC measures for technical data provided by the field staff, including field measurement data;
- Adherence to work schedules provided by the PM
- Identification of problems at the field team level, resolving difficulties in consultation with the PM, implementing and documenting corrective action procedures and providing communication between field sampling teams, PM, and ECB Laboratory
- Participation in preparation of the draft and final reports
- Input related to schedule and quality issues
- Preparation of DQCRs for the project, maintenance of copies of these documents at the site, and submittal of these documents to the PM on a weekly basis (if no problems occur at the site) or on a daily basis (if problems occur at the site) until the problems have been corrected.

5.6.2 Drillers

The drillers are responsible for drilling, operation and maintenance of the drilling equipment and obtaining all samples as required. A geologist or geological engineer will be the Field Team Leader for all activities involving drill crew operations. The drillers will advise the Field Team Leader on the best way to achieve the goals of the drilling program and direct the work of the driller's helpers.

5.6.3 Site Safety and Health Officer

The Site Safety and Health Officer (SSHO) will be responsible for:

- Updating equipment or procedures based on information gathered during the site inspection,
- Modifying the levels of protection based on site conditions,
- Determining and posting locations and routes to medical facilities and arranging for emergency transportation to medical facilities,

- Notifying local public emergency officers including police and fire departments of the nature of the team's operations and posting their telephone numbers,
- Examining work party members for symptoms of exposure or stress,
- Providing emergency medical care and first aid as necessary.

5.7 DATA VALIDATION

Data validation will be performed in accordance with requirements defined in the SSA. In general, data validation will be performed on at least 10% of the analyses by media and data review will be performed on 100% of the analyses by the Project Chemist.

Data validation will be performed on laboratory data for organic, inorganic and water quality parameter analyses to assess the quality and usability of the data. During validation, the following will be evaluated at a minimum:

- sample receipt,
- COC.
- sample documentation,
- sample holding times,
- initial and continuing instrument calibrations,
- gas chromatograph/mass spectrometer (GC/MS) tuning information,
- surrogate analyses,
- blank analyses (laboratory, field/rinsate, trip and preparation),
- MS/MSD analyses,
- LCS analyses,
- duplicate analyses (laboratory and field),
- internal standard performance,
- dilution and calculation discussions, and
- actions taken.

The evaluation will also include a statement of data quality that discusses the usability of the data.

Sites being investigated under the RAMS Program usually are not initially subject to the requirements of the CERCLA but at some point in time may eventually become subject to CERCLA and under the jurisdiction of the USEPA. Therefore, the evaluation will be based upon the *National Functional Guidelines for Organic Data Review* (U.S. EPA, February 1994), *National Functional Guidelines for Inorganic Data Review* (USEPA, February 1994) and *Minimum Chemistry Data Reporting Requirements for DERP and Superfund Projects* (USACE, August 1989).

5.8 REPORTING RESPONSIBILITY

The project team will be responsible for the preparation of the report.

6. LABORATORY ANALYTICAL AND PREPERATION PROCEDURES

This project will include the analysis of surface and subsurface soil, and sediment samples for volatile organic compounds (VOCs), selected Metals, and Cyanide (CN). It will also include the analysis of surface water and ground-water samples for VOCs, selected Metals, and Cyanide. Laboratory analysis of Water Quality Parameters/Design Parameters (which includes common ions) for surface water and ground-water samples may include some or all of the following: Alkalinity, Hardness (as calcium carbonate [CaCO₃]), Conductivity, Total Dissolved Solids (TDS), Total Organic Carbon (TOC), Sodium Adsorption Ratio (calculated), Nitrate/Nitrite, Sulfate, Carbonate, Bicarbonate, Chloride and Ammonia. Both Total and Dissolved Metals analyses may be performed on surface water and ground-water samples. A series of soil and sediment leaching procedures may be performed on these samples (see table 6-1 and section 6.2). Some other analyses may be performed on soil samples are given in table 6-1. All samples will be shipped to the ECB Laboratory for analyses (please see Section 3.7 for shipping address). The CSM may be used for specialty tests. The analytical methods are presented in Table 6-1 and a summary of internal QC checks is presented in Table 6-2. Specific analytes included in each analyte group and their LRLs, MDLs and QC Limits are presented in Tables 6-3 through 6-7 at the end of this section. Descriptions of internal QC checks are provided below.

6.1 LABORATORY ANALYTICAL METHODS

The analytical methods to be utilized by the ECB Laboratory and/or by CSM are summarized in the following subsections.

6.1.1 Volatile Organic Compounds (VOCs)

Surface soil, subsurface soil, and ground-water samples for VOCs will be analyzed by Third Edition SW-846 Final Update III (SW-846) 8260. Method 8260 is based upon a purge-and-trap, gas chromatographic/mass spectrometric (GC/MS) procedure.

6.1.2 <u>Metals</u>

In general, surface soil, subsurface soil, sediment samples, surface water, and ground-water samples will be analyzed for selected Metals by SW-846 6010. Method 6010 is based on the simultaneous or sequential, multi-elemental determination of elements by ICP-AES. ICP measures element-emitted spectra by optical spectrometry. Surface water and ground-water samples will analyzed for total recoverable and dissolved (Total and Dissolved) metals to assess the extent and potential for transport of metals through the surface water and ground water.

The soil and sediment samples will be digested by SW-846 3050A, while water samples will be digested using SW-846 3005A, prior to analysis.

6.1.3 Cyanide (CN)

Surface soil, subsurface soil, sediment samples, surface water, and ground-water samples will be analyzed for Cyanide by SW-846 9012 or by the Standard Method 4500. Total cyanide analysis is based on the decomposition of nearly all cyanides in the presence of strong acid, magnesium chloride, and heat during a one hr reflux distillation. The released cyanide in the form of hydrogen-cyanide (HCN) is captured in a sodium hydroxide (NaOH) trap where it is analyzed. The determination of Weak Acid Dissociable (WAD) Cyanide is based on the pretreatment of the sample with a week acid. Hydrogen cyanide is liberated from a slightly acidified sample (pH 4.5 to 6.0). The method does not recover the tightly bound cyanide complexes. The WAD cyanides that are captured in the NaOH trap are analyzed by a method the same as for total cyanide analysis. In many instances, stakeholders may prefer WAD Cyanide analysis that is generally considered to be more conservative with more reliable reported values. Stakeholders will be consulted for preferred method.

6.1.4 Water Quality Parameters

Surface water and ground-water samples will be prepared and analyzed for the determination of some or all of the following:

- Alkalinity using EPA 310.2,
- Hardness (as CaCO₃) calculated using ASTM 2340D and the total calcium/magnesium results,
- Conductivity using SW-846 9050A, 1985,
- TDS using EPA 160.1,
- Nitrate/Nitrite as N using EPA 353.2,
- Sulfate using EPA 375.2,
- Carbonate using EPA 310.2,
- Bicarbonate using EPA 310.2,
- Chloride using EPA 325.2,
- Fluoride using EPA 340.1, 340.2 or 340.3,
- Ammonia using EPA 350.1.

6.1.5 Soil Analyses (misc)

Several miscellaneous soil analyses that may be used are given in Table 6.1.

6.1.6 Soils Geotechnical Tests

Various geotechnical tests that may be used are available in Appendix C.

6.1.6.1 Water Holding Capacity (WHC)

Water Holding Capacity is a measure of the amount of water can be contained in the soil and is directly related to the quantity of micropores within the soil, thus is influenced by the texture, structure, and organic matter content. The WHC is determined using a method adopted from the American Society of Agronomy, Methods of Soil Analysis, Part 1 - Physical and Mineralogical Properties.

6.2 SOLID/SEDIMENT AND LEACHATE SAMPLE PREPERATION PROCEDURES

Soils and rocks from the mine waste piles are collected and analyzed by two methods. The solid sample (grab sample or composite) are screened through a $^{1}/_{4}$ inch stainless steel sieve, then placed in pre-cleaned 125 mL glass jars. Sediment samples are obtained from the stream and placed in sample containers. See Table 7-1 and 7-2 for sample container, preservation, and holding time requirements. Specific sampling and analysis procedures will be given in the SSA to the Work Plan. Portions of the samples are sent to the laboratory for the following analyses.

- A portion of the samples is sent for solid sample analysis as is given in section 6.1 above.
- A portion is sent to the lab for leachate analysis. The analytical method is given above in section 6.1. The leachate preparation is as follows:

6.2.1 Mine Waste Pile Soils for Leachate Preparation.

6.2.1.1 Colorado Department of Minerals and Geology (DMG) Procedure

This extraction consists of combining a 2 : 1 mixture of deionized water and solid, and let set for a set period time. The filtered extract is then analyzed for metals.

The deionized leachate is collected as follows: 150 mL (dry volume) of the prescreened soil is placed in a 1-liter plastic beaker along with 300 mL of deionized water. This is vigorously mixed for 15 seconds; the top is wrapped in plastic, then let settle for 90 minutes. This time is the time it will generally take for the soils fraction to settle out. The liquid is then filtered through a fine grade soil filter (approximately 2 micron). A portion of the liquid is analyzed for total acidity, pH, and specific conductance. The remaining liquid sample is acidified with nitric acid and analyzed for selected total metals.

6.2.1.2 Meteoric Water Mobility Procedure (MWMP)

The purpose of MWMP is to evaluate the potential for dissolution and mobility of certain constituents from a mine rock sample by Meteoric water. The method is based upon data collected for wet

depositions in Nevada, obtained from Desert Research Institute (DRI), University of Nevada Systems, Type II reagent water mostly simulates meteoric water in Nevada in terms of both composition and pH range, Meteoric Water Mobility Procedures, 9/19/90, and also Standard Methods for the Examination of Water and Wastewater, 1992.

6.2.2 Stream Sediment for Leachate Preparation.

Sediment samples are collected and analyzed by two methods. The sediment samples are analyzed as a solid by method as given in section 6.1. The leachate obtained from the sediment samples is analyzed for selected metals as shown in section 6.1. The leachate can be obtained by methods from mild dissolution to aggressive digestion on sediment of <80-mesh. The leachate methods are as follows:

6.2.2.1 DMG Procedure

This extraction is a 2:1 by volume deionized water extraction process as is used for the waste rock analysis in section 6.2.1.

6.2.2.2 Toxicity Characteristic Leaching Procedure (TCLP) (Modified EPA Method 1311)

The TCLP is a standard procedure used by the EPA to classify solid materials as hazardous wastes as designated for landfill disposal. The filtered leachate is then analyzed by method 6010.

The extraction solution is prepared by adding 5.7 mL of glacial acetic acid to 500 mL of water. To this is added 64.3 mL of 1 N NaOH and the volume is brought to 1 liter with water. The final solution should be pH 5. 2 grams of < 800-mesh sediment is added to 40 mL of the extraction solution in a 125 mL bottle. The bottle is agitated end over end using a rotary tumbler for 24 hrs. At the end of the 24 hrs the filtrate is filtered through a 0.45 micron filter, acidified with nitric acid and analyzed using ICP-AES.

6.2.2.3 Modified Synthetic Precipitation Leaching Procedures (SPLP) (EPA Method 1312).

The SPLP, in this unmodified form, is used by the USEPA to determine the potential mobility metals in a solid if exposed to rain or snowmelt and accounts to some extent for effects of acid rain.

The extraction fluid is prepared by adding a 60/40 weight percent mixture of sulfuric acid and nitric acid to reagent water until the pH is 5.00 +/- 0.05. 2 grams of < 80-mesh sediment is added to a 125 mL bottle containing 40 mL of the extraction solution. The bottle is then agitated end over end using a rotary tumbler for 18 hours. At the end of the 18 hrs the filtrate is filtered through a 0.45 micron filter, acidified with nitric acid and analyzed using ICP-AES.

6.2.2.4 Hydrogen Peroxide (H_2O_2) Acid Producing Potential (APP) Test (EPA Field and Lab Methods 3.2.11).

The H_2O_2 APP Test is a procedure that can be used to predict if a mine waste material will be an acid producer or a neutralizer. The APP releases sulfide material and the extraction fluid will indicate what metal can be found in a solid.

1 gram of < mesh sediment is placed in a Teflon beaker, 12 mL of 30 % hydrogen peroxide is added and heated to 40° C. The beaker is removed from the heat and the reaction is allowed to go to completion as is shown when bubbling has ceased. An additional 6 mL of 30 % hydrogen peroxide is added to the beaker and the reaction is once again allowed to go to completion. The beaker is then heated to 90°C and allowed to react until any unreacted hydrogen peroxide in the beaker is destroyed. The beaker is not allowed to go to dryness. The sides of the beaker is then washed with 50 mL of distilled water, and then heated to boiling to drive of any dissolved carbon dioxide. The beaker is then cooled to room temperature and titrated with 0.0100 N NaOH, that is free of carbon dioxide, to a pH of 7.0 using a pH meter. The liquid is filtered through a 0.45 micron filter, acidified with nitric acid and analyzed using ICP-AES, Hageman, Phillip L, and Briggs, Paul H, 2000.

6.2.2.5 Amorphous Digestion

An amorphous digestion is carried out in order to determine if iron and manganese oxides are present in the sediments analyzed. Like the APP, this procedure tests for specific species within the sediment.

The extraction solution is prepared that contains equal volumes of $0.2 \text{ M} \text{ (NH}_4)_2\text{C}_2\text{O}_4$, $0.2 \text{ M} \text{ H}_2\text{C}_2\text{O}_4$ $2\text{H}_2\text{O}$, and 0.1 M Ascorbic Acid. 80 mL of the extraction solution is added to 2 grams of <80-mesh sediment in a teflon beaker. The beaker is heated to 90° C for 30 minutes. Samples are filtered with a 0.45 micron filter, acidified with nitric acid, and analyzed using ICP-AES, Duggan, Laura, Colorado School of Mines, Masters Thesis, 1993.

6.3 ACID BASE ACCOUNTING

Acid base accounting is a static procedure that can be used to predict if a sample mine overburden material will be an acid producer or a neutralizer. The acid producing potential in a rock is tied directly to the amount of sulfides bound up in the rock in various forms. When these sulfides minerals are exposed to water and air, the sulfur is oxidized to form more soluble oxygen containing anions resulting in the release of the metallic ions and acids. The lower pH can then result in additional metals release. Acid Base Accounting is an analytical test that can determine the potential of the sample to produce acid and release metallic ions. The analysis will be performed as the Acid Base Accounting which included acid-base potential, acid potential, neutralization potential, and sulfur forms (sulfide, sulfite, sulfite etc.).

SMP buffer, which is a measure of the buffering capacity and exchangeable acidity can also be performed. See Table 6-1.

6.4 GROUND WATER AND SURFACE WATER SAMPLES

Water samples are obtained from monitoring wells and streams and sent to the laboratory for analysis. See Table 7-2 for sample container, preservation, and holding time requirements. Specific sampling procedures and required analyses are given in the Site Specific Addendum (SSA) to the Work Plan Portions.

6.5 LABORATORY QC SAMPLES

Laboratory QC samples will be included in the analyses as are given in this section.

6.5.1 Laboratory Blank Samples

Blank samples are analyzed to check for potential contamination by constituents derived from a source other than the sample (i.e., sample handling or laboratory procedures). Method blank samples are deionized, distilled water that are prepared and analyzed as a sample. The detection of contaminants in the method blank indicates potential contamination from laboratory sources (i.e., contaminated reagents, cleaned laboratory equipment or persistent contamination due to the presence of contaminants in the ambient laboratory air). A method blank will be prepared and analyzed with each batch of 20 or fewer samples. If analytes are detected above the limits listed on Table 6-2, preparation and analysis will be stopped and the source of contamination found, documented and eliminated. All samples following the last contaminant-free method blank that contained the contaminant will be re-prepared and re-analyzed. Other blank samples (such as preparation blanks, instrument blanks, calibration blanks, etc.) will be run as required by specific methods.

6.5.2 Surrogate Standards for Organic Analysis

A surrogate is a compound that is chemically similar to the compounds of interest in the field or unknown sample, but which is not normally found in environmental samples. Surrogates are added to all blanks, samples and spiked samples prior to preparation and analysis. The surrogate percent recovery is a measure of the effectiveness of the preparation and analysis methods on the field or unknown sample. The surrogate recovery must fall within the accuracy limits established on Tables 6-2. Any sample with a surrogate recovery that exceeds the QC limits must be re-prepared and re-analyzed in accordance with the Laboratory's LQMM, with both sets of data reported unless otherwise approved by the Project Chemist.

6.5.3 Internal Standards

An internal standard compound (the peak of which is well separated from the peaks of the components of the sample, but close to the peak of the sample components) is introduced into each sample and standard. The ratio of analyte to internal standard peak area serves as the analytical parameter measurement. Internal standards ensure that Gas Chromatography/Mass Spectroscopy (GC/MS) sensitivity and response are stable during each analysis for VOCs. Internal standard counts must not vary by more than a factor of two (-50% to 100%) from the associated calibration standard and the retention time of the internal standard must not vary more than +/- 30 seconds from the retention time of the associated calibration standard. Samples with internal standards outside these limits must be reanalyzed.

6.5.4 Matrix Spike/Matrix Spike Duplicates (MS/MSD) - Organics

An MS sample is an aliquot of the environmental sample that has been spiked with known concentrations of analytes. The MS sample is prepared and analyzed by the same method as the unspiked field sample. An MSD sample is a duplicate of the MS, which is prepared and analyzed as described above. The percent recovery is used to evaluate the effect the sample matrix has on analytical bias. The RPD of the spike duplicates is calculated and used to assess the analytical precision. Limits for percent recovery/RPD are listed on the Tables at the end of this section. No action will be taken to re-extract or re-analyze samples if MS/MSD precision and/or accuracy results are outside these limits unless other quality control parameters so merit.

6.5.5 Matrix Spike/Matrix Spike Duplicates - Inorganics

If the spike recovery results exceed the limits on Tables 6-2, re-preparation and/or re-analysis will be performed as necessary to confirm matrix interference. If matrix interference is confirmed, the respective sample result will be qualified during data validation. For example, if the spike recovery result for an analyte is below the value listed on Tables 6-2, but above thirty percent (30%), the result will be qualified as estimated with a "J" (estimated) code. If the spike recovery result for an analyte is less than thirty percent (30%) and the respective sample result is reported as not detected at (or less than) the reporting limit, the associated samples will be qualified with an unusable code ("R").

In instances where the sample concentration exceeds the spike concentration by a factor of four (4) or more, the limits on Table 6-2 may not apply and professional judgment will be used as to application of acceptance limits. However, no more than twenty percent (20%) of all spike recovery data reported for a given analyte may be reported where the sample concentration exceeds the spike concentration by a factor of four (4) or more. If the frequency exceeds twenty percent (20%), the MS and MSD samples will be prepared again at a higher spike concentration.

6.5.6 <u>Laboratory Control Samples/Laboratory Control Sample Duplicates</u>

LCS/ LCSD are samples which have been prepared in a matrix similar to that of the field samples and which have been spiked with known concentrations of analytes. The LCS/LCSD standard is prepared independently of calibration standards to provide a check of instrument calibration. The LCS and LCSD are prepared and analyzed by the same methods as the field sample. The percent recovery of the analytes in the LCS and LCSD indicate whether the analytical method is within established control limits. Control limits are presented on Table 62. If the control limits are exceeded, appropriate corrective actions will be taken in accordance with ECB Laboratory's LQMM.

6.6 FIELD QC SAMPLES

During the field activities for this project, QC samples will be collected/generated to assess the quality of the data resulting from the field sampling program and the analytical laboratory procedures. QC samples will consist of blanks (trip blanks, potable water supply blank, and/or source water supply blank), field duplicate samples and site-specific MS/MSD samples that will be sent to ECB Laboratory.

6.6.1 Blank Samples

Field blank samples are samples taken in the field of sources used in the field or samples to determine if any contamination has been put into the samples on transit.

Trip blanks samples will be submitted and analyzed to assess the quality of the data resulting from the field sampling program. Trip blanks will be used to assess the potential for VOC contamination of water samples due to contaminant migration during sample collection and handling, shipment and storage procedures. One VOC trip blank with deionized distilled ultra pure water will be included with each shipment of aqueous VOC samples. Trip blanks will be analyzed for all VOC analytes of concern. The objective is for the trip blanks to contain no detectable analytes above the analytical reporting limits and for all samples to be properly preserved when received at the laboratory.

Trip blanks will consist of containers (VOC vials) of reagent-grade deionized distilled ultra pure water filled in the field with the source water from Laboratory. Several 5-liter containers of trip blank source water obtained from ECB Laboratory will accompany the VOC containers (VOC vials) from the lab to the field. Trip blanks will be kept with the aqueous field samples for VOCs analysis and shipped with them to the lab for analysis. The source water for the trip blanks will also be used for the final rinse in the decontamination procedure.

6.6.2 Field Duplicate Samples

Field duplicate samples will be analyzed to assess the quality of the data resulting from the field sampling program. Field duplicate samples will be analyzed to provide an indication of precision with respect to sample collection procedures and analytical reproducibility. Field duplicates are samples taken in the field at the same time and in the same manner as the original sample for the purpose of verifying the sampling technique. Field duplicate soil samples will be prepared by mixing the original soil sample and dividing it into two subsamples, except for VOC analyses. Field duplicate soil samples requiring VOCs analyses will consist of separate grab samples taken from the same interval. The general level of the QC effort will be one field duplicate for every 10 primary samples. Three times the standard volume of sample is required at locations where QC and MS/MSD samples are collected.

Field duplicates (D1 and D2) are compared using the following RPD calculation:

$$RPD = [D1 - D2/(D1 + D2/2)] \times 100$$

where: D1 is the measured concentration of the first sample aliquot

D2 is the measured concentration of the second sample aliquot

6.6.3 Matrix Spike/Matrix Spike Duplicate Samples

Matrix spikes provide information about the effect of the sample matrix on the digestion and measurement methodology; all matrix spikes will be performed in duplicate and are referred to as MS/MSD samples. MS/MSD samples are QC samples. Site-specific MS/MSD samples will be collected at a frequency of one per twenty (20) or fewer primary samples per sample matrix (i.e., surface soil, subsurface soil, ground water, etc.). Three times the standard volume of sample is required at locations where QC and MS/MSD samples are collected.

Table 6-1
ANALYTICAL METHODS

| Analyte | Soil Method | Water Method | | |
|--|--|--|--|--|
| Volatile Organics ^a | 8260 | 8260 | | |
| Inorganics Metals/ (Metals) ^a | 1 | 1 | | |
| Aluminum (Al) | 3050A/6010 | 3005A/6010 | | |
| Antimony (Sb) | 3050A/6010 | 3005A/6010 | | |
| Arsenic (As) | 3050A/6010 | 3005A/6010 | | |
| Barium (Ba) | 3050A/6010 | 3005A/6010 | | |
| Beryllium (Be) | 3050A/6010 | 3005A/6010 | | |
| Cadmium (Cd) | 3050A/6010 | 3005A/6010 | | |
| Calcium (Ca) | 3050A/6010 | 3005A/6010 | | |
| Chromium (Cr) | 3050A/6010 | 3005A/6010 | | |
| Cobalt (Co) | 3050A/6010 | 3005A/6010 | | |
| Copper (Cu) | 3050A/6010 | 3005A/6010 | | |
| Iron (Fe) | 3050A/6010 | 3005A/6010 | | |
| Lead (Pb) | 3050A/6010 | 3005A/6010 | | |
| Magnesium (Mg) | 3050A/6010 | 3005A/6010 | | |
| Manganese (Mn) | 3050A/6010 | 3005A/6010 | | |
| Mercury (Hg) | 7471A | 7470A | | |
| Nickel (Ni) | 3050A/6010 | 3005A/6010 | | |
| Potassium (K) | 3050A/6010 | 3005A/6010 | | |
| Selenium (Se) | 3050A/6010 | 3005A/6010 | | |
| Silver (Ag) | 3050A/6010 | 3005A/6010 | | |
| Sodium (Na) | 3050A/6010 | 3005A/6010 | | |
| Thallium (Tl) | 3050A/6010 | 3005A/6010 | | |
| Vanadium (V) | 3050A/6010 | 3005A/6010 | | |
| Zinc (Zn) | 3050A/6010 | 3005A/6010 | | |
| Infrequently Analyzed Metals: Bismuth, Boron, Gallium, Lithium, Molybdenum, Phosphorus, Scandium, Strontium, Tin, Titanium | 3050A/6010 (This method is usable with some possible modifications) | 3005A/6010 (This method is usable with some possible modifications) | | |

| Analyte | Soil Method | Water Method |
|---|--|--|
| Inorganic Analytes | | |
| Cyanide (CN) | Standard Method 4500 or SW846 Method 9012 | Standard Method 4500 or Sw846 Method 9012 |
| Cyanide (WAD CN)* | Standard Method 4500 or SW846 Method 9012 | Standard Method 4500 or Sw846 Method 9012 |
| Water Quality Parameters/Design Parameter | ers | |
| Alkalinity (total, reported as carbonate and bicarbonate) | NA | EPA 310.2 |
| Hardness (as CaCO ₃) | NA | ASTM 2340D |
| Sodium Adsorption Ratio (calculated) | NA | Bohn et al ** |
| TDS | NA | EPA 160.1 |
| Nitrate/Nitrite | NA | EPA 353.2 |
| Sulfate | NA | EPA 375.2 |
| Carbonate | NA | EPA 310.2 |
| Bicarbonate | NA | EPA 310.2 |
| Chloride | NA | EPA 325.2 |
| Fluoride | 340.1, 340.2, 340.3 | 340.1, 340.2, 340.3 |
| Ammonia | NA | EPA 350.1 |
| Conductivity | 9050A | 9050A |
| TOC | 9060 | 9060 |
| Leaching Procedures | | |
| DMG Procedure | Section 6.2.1.1 | |
| TCLP | Modified EPA 1311 | |
| SPLP | EPA Method 1312 | |
| H ₂ O ₂ APP | EPA Method 3.2.11 | |
| Amorphous Digestion | Section 6.2.2.5 | |
| Meteoric Water Mobility Procedure | Section 6.2.1.2 | |
| Other Soil Analyses | 1 | |
| Soil Conductivity | ASTM D1125 | |
| Acid-Base Accounting | *** | |
| SMP Buffering Capacity | *** | |
| Cation Exchange Capacity | *** | |
| Soil Water Holding Capacity (WHC) | Section 6.1.6.1 | |

Notes:

a = EPA SW - 846 Methods

NA = Not Applicable * = WAD - Weak Acid Dissociable

^{** =} Bohn et al in Soil Chemistry, 1985 (calculated as described using Ca, Mg, K, Na results).

*** = Nevada Bureau of Mining Regulations and Reclamation, Waste Rock and Overburden Evaluation, Sept 14, 1990.

Table 6-2
INTERNAL QUALITY CONTROL CHECKS

| Analytic Group | La | ıb Blank | F | ield Split | LC | S/LCSD | Surrog | ate Spike | Inte Stand | | М | S/MSD | Tri | ip Blank |
|-------------------------------------|------|---|------|---------------------------------------|------|-------------------------------|-----------------|-------------------------------|-----------------|---------|-------|-------------------------------|-----|----------------------|
| | F | Limits | F | Limits | F | Limits | F | Limits | F | Limits | F | Limits | F | Limits |
| Volatile Organics | 1/20 | <pql<sup>a</pql<sup> | 1/10 | 50% RPD- Soil 30% RPD- Water | 1/20 | Table 6-3 and Table 6-4 | Every Sample | Table 6-3 and Table 6-4 | Every Sample | Section | 1/20 | Table 6-3 and Table 6-4 | * | <pql<sup>a</pql<sup> |
| Total Cyanide and WAD Cyanide | 1/20 | < RL | 1/10 | 50% RPD- Soil 30% RPD- Water | 1/20 | Table 6-7 | Every Sample | Table 6-7 | QC | | 1 /20 | Table 6-7 | NA | NA |
| Metals (soil) Metals (water) | 1/20 | <rl< td=""><td>1/10</td><td>50% RPD- Soil 30% RPD- Water</td><td>1/20</td><td>Table 6-5 and Table 6-6</td><td>NA</td><td>NA</td><td>NA</td><td>NA</td><td>1/20</td><td>Table 6-5 and Table 6-6</td><td>NA</td><td>NA</td></rl<> | 1/10 | 50% RPD- Soil 30% RPD- Water | 1/20 | Table 6-5 and Table 6-6 | NA | NA | NA | NA | 1/20 | Table 6-5 and Table 6-6 | NA | NA |

Notes:

PQL = Practical Quantitation Limit

RL = Reporting Limit

NA = Not Applicable

-- = Not available

F = Frequency

a = Lab solvents include acetone, methylene chloride, 2-butanone and toluene

b = Common Phthalate esters

* = One trip blank per every cooler containing water volatile organic samples

QC - Cyanide analysis requires an internal calibration curve and a midlevel continuing calibration

Table 6-3

Volatile Organic Compounds
Reporting and QC Limits
for Soil Samples
Method 8260

| CAS No. | Analyte | MDL (µg/kg) | LRL (µg/kg) | Acceptance Range * (% Recovery) | RPD (%) |
|------------|---------------------------|-------------|-------------|---------------------------------------|------------|
| 100-41-4 | Ethylbenzene | 1 | 5.0 | 60-140 | 25 |
| 100-42-5 | Styrene | 1 | 5.0 | 60-140 | 25 |
| 10061-01-5 | cis-1,3-Dichloropropene | 1 | 5.0 | 60-120 | 25 |
| 10061-02-6 | trans-1,3-Dichloropropene | 1 | 5.0 | 50-150 | 25 |
| 107-06-2 | 1,2-Dichloroethane | 1 | 5.0 | 60-140 | 25 |
| 108-05-4 | Vinyl Acetate | 20 | 100 | 50-150 | 25 |
| 108-10-1 | 4-Methyl-2-pentanone | 20 | 50 | 50-150 | 25 |
| 108-88-3 | Toluene | 1 | 5.0 | 59-139 | 25 |
| 108-90-7 | Chlorobenzene | 1 | 5.0 | 60-133 | 25 |
| 124-48-1 | Dibromochloromethane | 1 | 5.0 | 60-140 | 25 |
| 127-18-4 | Tetrachloroethene | 1 | 5.0 | 60-140 | 25 |
| 1330-20-7B | meta-/para-Xylenes | 1 | 5.0 | 60-140 | 25 |
| 156-59-2 | cis-1,2-Dichloroethene | 1 | 5.0 | 60-140 | 25 |
| 156-60-5 | trans-1,2-Dichloroethene | 1 | 5.0 | 60-140 | 25 |
| 56-23-5 | Carbon Tetrachloride | 1 | 5.0 | 60-140 | 25 |
| 591-78-6 | 2-Hexanone | 20 | 50 | 60-140 | 25 |
| 67-64-1 | Acetone | 20 | 50 | 50-150 | 25 |
| 67-66-3 | Chloroform | 1 | 5.0 | 60-140 | 25 |
| 71-43-2 | Benzene | 1 | 5.0 | 66-142 | 25 |
| 71-55-6 | 1,1,1-Trichloroethane | 1 | 5.0 | 60-140 | 25 |
| 74-83-9 | Bromomethane | 1 | 5.0 | 50-150 | 25 |
| 74-87-3 | Chloromethane | 1 | 5.0 | 50-150 | 25 |

| CAS No. | Analyte | MDL (μg/kg) | LRL (µg/kg) | Acceptance Range * (% Recovery) | RPD (%) |
|------------|---------------------------|-------------|-------------|---------------------------------------|---------|
| 75-00-3 | Chloroethane | 1 | 10 | 50-150 | 25 |
| 75-01-4 | Vinyl Chloride | 1 | 5.0 | 50-150 | 25 |
| 75-09-2 | Methylene Chloride | 1 | 5.0 | 60-140 | 25 |
| 75-15-0 | Carbon Disulfide | 1 | 5.0 | 60-140 | 25 |
| 75-25-2 | Bromoform | 1 | 5.0 | 60-140 | 25 |
| 75-27-4 | Bromodichloromethane | 1 | 5.0 | 60-140 | 25 |
| 75-34-3 | 1,1-Dichloroethane | 1 | 5.0 | 60-140 | 25 |
| 75-35-4 | 1,1-Dichloroethene | 1 | 5.0 | 59-172 | 25 |
| 78-87-5 | 1,2-Dichloropropane | 1 | 5.0 | 60-140 | 25 |
| 78-93-3 | 2-Butanone | 20 | 50 | 50-140 | 25 |
| 79-00-5 | 1,1,2-Tric hloroethane | 1 | 5.0 | 60-140 | 25 |
| 79-01-6 | Trichloroethene | 1 | 5.0 | 62-137 | 25 |
| 79-34-5 | 1,1,2,2-Tetrachloroethane | 1 | 5.0 | 60-140 | 25 |
| 95-47-6 | ortho-Xylene | 1 | 5.0 | 60-140 | 25 |
| CAS No. | Surrogates | | | Acceptance | RPD |
| | | | | Ranges (%) | (%) |
| 17060-07-0 | 1,2-Dichloroethane-d4 | | | 70-121 | 25 |
| 1868-53-7 | Dibromofluoromethane | | | 80-120 | 25 |
| 2037-26-5 | Toluene-d8 | | | 81-117 | 25 |
| 460-00-4 | 4-Bromofluorobenzene | | | 74-121 | 25 |

^{*} Acceptable ranges are compound specific for the LCS/Spike Duplicate and MS/MSD

Table 6-4

Volatile Organic Compounds

Reporting and QC Limits

for Water Samples

Method 8260

| CAS No. | Analyte | MDL (μg/L) | LRL (µg/L) | Acceptance Range * | RPD (%) |
|------------|---------------------------|---------------|------------|-----------------------|---------|
| | | (Fg/ =) | (FB/ 22) | (% Recovery) | (,,, |
| 100-41-4 | Ethylbenzene | 0.5 | 2.0 | 70-130 | 25 |
| 100-42-5 | Styrene | 0.5 | 2.0 | 70-130 | 25 |
| 10061-01-5 | cis-1,3-Dichloropropene | 0.5 | 2.0 | 70-130 | 25 |
| 10061-02-6 | trans-1,3-Dichloropropene | 0.5 | 2.0 | 60-140 | 25 |
| 107-06-2 | 1,2-Dichloroethane | 0.5 | 2.0 | 70-130 | 25 |
| 108-05-4 | Vinyl Acetate | 10 | 50 | 60-140 | 25 |
| 108-10-1 | 4-Methyl-2-pentanone | 10 | 25 | 60-140 | 25 |
| 108-88-3 | Toluene | 0.5 | 2.0 | 76-125 | 25 |
| 108-90-7 | Chlorobenzene | 0.5 | 2.0 | 75-130 | 25 |
| 124-48-1 | Dibromochloromethane | 0.5 | 2.0 | 70-130 | 25 |
| 127-18-4 | Tetrachloroethene | 0.5 | 2.0 | 70-130 | 25 |
| 1330-20-7B | meta-/para-Xylenes | 0.5 | 2.0 | 70-130 | 25 |
| 156-59-2 | cis-1,2-Dichloroethene | 0.5 | 2.0 | 70-130 | 25 |
| 156-60-5 | trans-1,2-Dichloroethene | 0.5 | 2.0 | 70-130 | 25 |
| 56-23-5 | Carbon Tetrachloride | 0.5 | 2.0 | 70-130 | 25 |
| 591-78-6 | 2-Hexanone | 10 | 25 | 70-130 | 25 |
| 67-64-1 | Acetone | 10 | 25 | 60-140 | 25 |
| 67-66-3 | Chloroform | 0.5 | 2.0 | 70-130 | 25 |
| 71-43-2 | Benzene | 0.5 | 2.0 | 76-127 | 25 |
| 71-55-6 | 1,1,1-Trichloroethane | 0.5 | 2.0 | 70-130 | 25 |
| 74-83-9 | Bromomethane | 0.5 | 2.0 | 60-140 | 25 |
| 74-87-3 | Chloromethane | 0.5 | 2.0 | 60-140 | 25 |

| CAS No. | Analyte | MDL (µg/L) | LRL (µg/L) | Acceptance Range * | RPD (%) |
|------------|---------------------------|------------|---------------|--------------------------------|---------|
| 75-00-3 | Chloroethane | 0.5 | 2.0 | (% Recovery) 60-140 | 25 |
| 75-01-4 | Vinyl Chloride | 0.5 | 2.0 | 60-140 | 25 |
| 75-09-2 | Methylene Chloride | 0.5 | 2.0 | 70-130 | 25 |
| 75-15-0 | Carbon Disulfide | 0.5 | 2.0 | 70-130 | 25 |
| 75-25-2 | Bromoform | 0.5 | 2.0 | 70-130 | 25 |
| 75-27-4 | Bromodichloromethane | 0.5 | 2.0 | 70-130 | 25 |
| 75-34-3 | 1,1-Dichloroethane | 0.5 | 2.0 | 70-130 | 25 |
| 75-35-4 | 1,1-Dichloroethene | 0.5 | 2.0 | 61-145 | 25 |
| 78-87-5 | 1,2-Dichloropropane | 0.5 | 2.0 | 70-130 | 25 |
| 78-93-3 | 2-Butanone | 10 | 25 | 60-140 | 25 |
| 79-00-5 | 1,1,2-Trichloroethane | 0.5 | 2.0 | 70-130 | 25 |
| 79-01-6 | Trichloroethene | 0.5 | 2.0 | 71-120 | 25 |
| 79-34-5 | 1,1,2,2-Tetrachloroethane | 1 | 2.0 | 70-130 | 25 |
| 95-47-6 | ortho-Xylene | 0.5 | 2.0 | 70-130 | 25 |
| CAS No. | Surrogates | | | Acceptance Ranges (%) | RPD (%) |
| 17060-07-0 | 1,2-Dichloroethane-d4 | | | 76-114 | 25 |
| 1868-53-7 | Dibromofluoromethane | | | 80-120 | 25 |
| 2037-26-5 | Toluene-d8 | | | 88-110 | 25 |
| 460-00-4 | 4-Bromofluorobenzene | | | 86-115 | 25 |

^{*} Acceptable ranges are compound specific for the LCS/Spike Duplicate and MS/MSD

Table 6-5

Selected Metals Reporting and QC Limits for Soil Samples ICP Trace

LCS, MS/MSD Acceptance Range: 80-120% RPD limit: 25%

| Analyte | Method Detection Limit (mg/Kg) | Laboratory Reporting Limit (mg/Kg) |
|------------------|--------------------------------|------------------------------------|
| Aluminum (Al) | 4 | 12 |
| Antimony (Sb) | 1.2 | 4 |
| Arsenic (As) | 0.6 | 3 |
| Barium (Ba) | 0.1 | 0.5 |
| Beryllium (Be) | 0.1 | 0.4 |
| Boron (B) | 0.8 | 4 |
| Cadmium (Cd) | 0.1 | 0.5 |
| Calcium (Ca) | 20 | 60 |
| Chromium (Cr) | 0.4 | 2 |
| Cobalt (Co) | 0.6 | 3 |
| Copper (Cu) | 0.4 | 2 |
| Iron (Fe) | 8 | 24 |
| Lead (Pb) | 0.4 | 2 |
| Magnesium (Mg) | 8 | 24 |
| Manganese (Mn) | 0.2 | 0.6 |
| Mercury (Hg) (a) | 0.002 | 0.01 |
| Molybdenum (Mo) | 0.4 | 2 |
| Nickel (Ni) | 0.6 | 2 |
| Potassium (K) | 20 | 60 |
| Selenium (Se) | 0.8 | 4 |
| Silver (Ag) | 0.2 | 1 |
| Sodium (Na) | 20 | 60 |
| Thallium (Tl) | 1 | 5 |

| Analyte | Method Detection Limit (mg/Kg) | Laboratory Reporting Limit (mg/Kg) | | |
|---|--------------------------------|------------------------------------|--|--|
| Titanium (Ti) | 0.2 | 1 | | |
| Vanadium (V) | 4 | 20 | | |
| Zinc (Zn) | 0.6 | 2 | | |
| Infrequently Analyzed Metals: Bismuth, Gallium, Lithium, Phosphorus, Scandium, Strontium, Tin | Method Criteria | Method Criteria | | |

⁽a) = Cold vapor method

Table 6-6

Selected Metals Reporting and QC Limits for Water Samples ICP Trace

LCS, MS/MSD Acceptance Range: 80-120% RPD limit: 25%

| Analyte | Method Detection Limit | Laboratory Reporting Limit |
|------------------|-------------------------------|-----------------------------------|
| | (μg/L) | (µg/L) |
| Aluminum (Al) | 20 | 60 |
| Antimony (Sb) | 3 | 15 |
| Arsenic (As) | 3 | 15 |
| Barium (Ba) | 0.5 | 2.5 |
| Beryllium (Be) | 0.5 | 2 |
| Boron (B) | 4 | 20 |
| Cadmium (Cd) | 0.5 | 2.5 |
| Calcium (Ca) | 100 | 300 |
| Chromium (Cr) | 2 | 10 |
| Cobalt (Co) | 3 | 15 |
| Copper (Cu) | 2 | 10 |
| Iron (Fe) | 40 | 120 |
| Lead (Pb) | 2 | 10 |
| Magnesium (Mg) | 40 | 120 |
| Manganese (Mn) | 1 | 3 |
| Mercury (Hg) (a) | 0.01 | 0.05 |
| Molybdenum (Mo) | 2 | 10 |
| Nickel (Ni) | 3 | 10 |
| Potassium (K) | 100 | 300 |
| Selenium (Se) | 4 | 20 |
| Silver (Ag) | 1 | 5 |
| Sodium (Na) | 200 | 400 |
| Thallium (Tl) | 5 | 25 |

| Analyte | Method Detection Limit (µg/L) | Laboratory Reporting Limit (µg/L) |
|---|-------------------------------|-----------------------------------|
| Titanium (Ti) | 1 | 5 |
| Vanadium (V) | 4 | 20 |
| Zinc (Zn) | 3 | 10 |
| Infrequently Analyzed Metals: Bismuth, Gallium, Lithium, Phosphorus, Scandium, Strontium, Tin | Method Criteria | Method Criteria |

a = Cold Vapor

Table 6-7

Selected Inorganic Methods Reporting and QC Limits (LCS and MS/MSD Acceptance Limits) for Soil and Water Samples

by Various Methods

| Analyte | Soil | Soil | Water | Water | LCS,MS,MSD | RPD |
|---|---------|---------|---------|---------|---------------------|-------|
| | MDL | LRL | MDL | LRL | Acceptance Range | Limit |
| | (mg/kg) | (mg/kg) | (mg/L) | (mg/L) | | |
| Alkalinity, EPA 310.2 | NA | NA | 7 | 20 | 80-120% | 20% |
| Hardness, ASPM 2340D | NA | NA | 7 | 21 | 80-120% | 25% |
| Total Dissolved Solids (TDS), EPA 160.1 | NA | NA | 5 | 15 | 80-120% | 20% |
| Nitrate/Nitrite, EPA 353.2 | NA | NA | 0.01 | 0.10 | 80-120% | 25% |
| Sulfate, EPA 375.2 | NA | NA | 6.0 | 20 | 80-120% | 25% |
| Carbonate, EPA 310.2 | NA | NA | 7 | 20 | 80-120% | 25% |
| Bic arbonate, EPA 310.2 | NA | NA | 7 | 20 | 80-120% | 25% |
| Chloride, EPA 325.2 | NA | NA | 0.5 | 2 | 80-120% | 25% |
| Fluoride, EPA 340.1, 340.2, or 340.3 | NA | NA | 0.5 | 2 | 80-120% | 25% |
| Ammonia, EPA 350.1 | NA | NA | 0.02 | 0.10 | 80-120% | 25% |
| Conductivity, 9050A and ASTM D1125 | NA | NA | NA | NA | NA | NA |
| Sodium Adsorption Ratio, by Bohn et al | NA | NA | 2 meq/L | 6 meq/L | 80-120% | 25% |
| Total and WAD Cyanide | 1 | 5 | 0.002 | 0.005 | 80-120 % | 20 % |

| Analyte | Soil | Soil | Water | Water | LCS,MS,MSD | RPD |
|--------------------|-------------|-------------|------------|------------|---------------------|-------|
| | MDL (mg/kg) | LRL (mg/kg) | MDL (mg/L) | LRL (mg/L) | Acceptance Range | Limit |
| | | | | | | |
| Soil pH, EPA 150.1 | NA | NA | NA | NA | NA | NA |

NA = Not Applicable

7. SAMPLE COLLECTION, HANDLING AND DOCUMENTATION PROCEDURES

Sample containers and preservatives will be obtained through the laboratory from vendors. All sample containers will be certified as clean by the vendor, and a certificate of analysis will be supplied with each lot ordered. These certificates will be kept on file by the Field Team Leader and a record of the bottle lot numbers and preservatives used in the field will be made in the field sampling log; bottle lots and preservatives will be traceable to the fieldwork event. Sample containers and preservatives to be used are listed in Tables 7-1 and 7-2 and are grouped by analytes and matrix. These tables also identify the sample holding times by analytical parameter.

In general, sample collection order will be from locations surrounding the areas of concern which are expected to contain the least amount of contamination, to the locations that may contain the most contaminated soils; this determination will be based on the topography of the area, with sampling locations at higher elevations sampled first. Up-gradient wells will be sampled first (again, based on the topography of the site).

The order of sample collection at each location and sample interval will consist of obtaining samples for VOC analyses followed by VOC samples for on-site VOC headspace analysis, then samples for Metals and Design Parameters or Water Quality Parameters/Design Parameters (when appropriate).

7.1 SAMPLING IDENTIFICATION SCHEME

Samples will be uniquely identified by using a system as outlined below. If a different nomenclature system is used it shall be outlined in the SSA.

UU-VVVV-XXXX-YY-ZZ

where:

- **UU** = Project Designation
- **VVVV** = Designation of Sampling Area Location and an alphabetical letter may be inserted at the end of the VVVV character designation for the sub-site identification
- **XXXX** = 4 Character Designation for Sample Type/Location, where the first two X-characters indicate sample type:

MW = Monitoring Well

SB = Soil Boring

SD = Sediment

SS = Surface Sample

SW = Surface Water

TB = Trip Blank

WB = Water Blank

The next two X-characters, 01 through 99, further define the sample location based on the boring or sampling site number. Trip blanks will be marked with a MW or SW designation to prevent preferential treatment by the laboratory.

- YY = 2 Character Designation for Sample Depth, using the bottom depth of the soil sample interval, such as using -04, for the 2 to 4-foot interval. Water samples (MW, TB, WB, and SW) will not require a YY designation.
- **ZZ** = 2 Character Designation for Samples, where:

01 = Normal Field Sample

02 = QC Duplicate

03 = Trip Blank

04 = Confirmation Sample

7.2 SAMPLE CUSTODY

Documentation of sample custody will be maintained for all samples, from the time the samples are collected until analytical reports are received. A sample is under custody if:

- It is in a person's possession
- It is in a person's view, after being in one's possession
- It was in a person's possession and that person placed it in a secure area

The sample collector and/or sample packer will be responsible for initiating and filling out the COC form. The COC form will be signed by the sample collector when the samples are relinquished to anyone else (this includes relinquishing samples to a designated sample packer). The field team member packaging the samples for overnight shipment will sign the COC (if different from the sample collector). It is not necessary for the overnight shippers to sign the COC; however the shipping bill will be retained by the designated sample packer for tracking purposes. A COC form will be completed daily for each cooler. The following information will be included on the COC:

- Sampler's signature and affiliation
- Project name unique to the investigated site and a unique LIMS # xxxxxx. A unique LIMS number will obtained from the USACE project Chemist for each site investigated.
- Date and time of collection
- Sample identification, soil boring or monitoring well identification
- Sample type, matrix
- Preservatives used
- Internal temperature of the shipping container upon receipt at the laboratory when the shipping container is opened
- Analyses requested
- Number of containers
- Dates, times, and signature(s) of person(s) relinquishing custody
- Dates, times, and signature(s) of person(s) accepting custody

- Method of shipment
- Special instructions and/or Remarks

Upon receipt at the laboratory, the person receiving and unpacking the samples will sign the COC and it will be kept with the project records. The USACE Project Chemist will be notified if any noncompliance to set sample criteria occurs. The person receiving and unpacking the samples will also fill out a Cooler Receipt Form for each cooler received at the laboratory. A copy of the Cooler Receipt Form will be provided to the USACE Project Chemist.

Please refer to Chapter 4 of ECB Laboratory's LQMM (Appendix D) and Work Plan SOP A13 for corresponding information. Examples of a COC Form and a Cooler Receipt Form are provided in ECB Laboratory's LQMM (Appendix D).

TABLE 7-1 SAMPLE CONTAINERS, PRESERVATION, AND HOLDING TIMES FOR SOIL SAMPLES

| Parameter | Container ^{1,2} | Preservation | Maximum Holding Times | |
|-------------------------|---|--------------|---------------------------------|---------------------------------|
| | | | Extraction | Analysis |
| Volatiles | 2 X 2 oz Glass, Teflon Septa, no headspace. | Ice to 4°C | NA | 14 days |
| Cyanide (total and WAD) | 1 x 4 oz Glass | Ice to 4°C | | 14 days |
| Metals ³ | 1 x 4 oz Glass | Ice to 4°C | 6 mo. (Mercury - 28 days) | 6 mo. (Mercury - 28 days) |
| Acid Base Accounting | 16 oz glass container or plastic bag container | | | NA ⁴ |
| TOC | 4 ox glass | Ice to 4°C | | ASAP |

¹ Three times the standard volume of sample is required at locations where QC and Matrix Spike/Matrix Spike Duplicate samples are collected.

- 2 Additional soil volume must be collected when leaching procedures are required.
- 3 Samples for metals need not be place in iced container.
- 4 If sample is kept in a closed to prevent contamination, no holding time is stated.

ASAP = As Soon As Possible NA = Not Applicable

TABLE 7-2 SAMPLE CONTAINERS, PRESERVATION, AND HOLDING TIMES FOR WATER SAMPLES

| Parameter | Container ¹ | Preservation | Maximum Holding Times: | | |
|---|---|---|------------------------|------------------------------------|--|
| | | | Extraction | Analysis | |
| Volatiles | 3 x 40 mL Glass, Septa vial, no headspace | Ice to 4°C HCl to pH <2 | NA | 14 days | |
| Dissolved Metals ² | 1 x 1 Liter Plastic | Preservative added after filtering by ECB Lab | NA | 6 months (Mercury - 28 days) | |
| Total Metals ² | 1 x 1 Liter Plastic | HNO ₃ to pH <2 Do not filter | NA | 6 months (Mercury - 28 days) | |
| Total and WAD Cyanide | 2 x 500 mL plastic | NaOH to pH > 12 | | 14 days | |
| Alkalinity | | | NA | 14 days | |
| Conductivity | | | NA | 28 days | |
| Sulfate | | | NA | 28 days | |
| Carbonate | 1 x 1 Liter Plastic | Ice to 4°C | NA | 28 days | |
| Bicarbonate | | | NA | 28 days | |
| Chloride | | | NA | 28 days | |
| Hardness | 1 x 250 mL Plastic | HNO ₃ to pH <2 Ice to 4°C | NA | 6 months | |
| Nitrate/Nitrite | 1 x 250 mL Plastic | H ₂ SO ₄ to pH <2 Ice to 4°C | NA | 28 days | |
| Sodium Adsorption Ratio (Ca, Mg, K, Na) | 1 x 1 Liter Plastic | HNO ₃ to pH <2 Do not filter | NA | 6 months | |
| Total Dissolved Solids (TDS) | 1 x 1 Liter Glass | Ice to 4°C To be filtered by ECB Lab | NA | 7 days | |
| Total Organic Carbon (TOC) | 1 x 1 Liter Glass | H ₂ SO ₄ or HCl to pH <2 Ice to 4°C | | 28 days, if preserved | |
| Ammonia | 1 x 500 mL Plastic | H ₂ SO ₄ to pH <2 Ice to 4°C | NA OG IMALI S | 28 days | |

¹ Three times the standard volume of sample is required at locations where QC and Matrix Spike/Matrix Spike Duplicate samples are collected.

NA = Not Applicable

² Samples for metals need not be place in iced container

8. PREVENTIVE MAINTENANCE PROCEDURES

8.1 FIELD EQUIPMENT

The field equipment for this project includes thermometers, turbidity meter, and multi-parameter probe [for measurement of pH, conductivity, oxidation reduction potential (ORP) or Eh, and dissolved oxygen (DO)]. Specific preventive maintenance procedures recommended by the manufacturer will be followed for field equipment.

Critical spare parts such as pH probes, electrodes and batteries will be kept on-site to minimize instrument down time. Backup instruments and equipment will be available on-site or within one-day shipment to avoid delaying field activities.

8.2 LABORATORY INSTRUMENTS

Routine preventive maintenance programs will be conducted by laboratory to minimize the occurrence of instrument failure and other system malfunctions. Section supervisors and/or analysts will perform routine scheduled maintenance and coordinate with the vendor for the repair of all instruments. All laboratory instruments will be maintained in accordance with manufacturer's specifications and the requirements of the specific method employed; this maintenance will be carried out on a regular, scheduled basis and will be documented in the laboratory instrument service logbook for each instrument. Emergency repair or scheduled manufacturers maintenance will be provided under a repair and maintenance contract with factory representatives. Lists of equipment available at the laboratory are available in the LQMM.

9. CALIBRATION PROCEDURES AND FREQUENCY

9.1 FIELD INSTRUMENTS

Maintenance and proper calibration of field instruments are essential to the field sampling program. Before mobilization to the field, all equipment will be checked for malfunctions and cleaned. In addition, all equipment will be calibrated by the sampling team to ensure proper working order and to ensure the integrity to the measured values. Calibration will be performed on a daily basis or more frequently as required. Calibration procedures provided by the manufacturer will be followed and are contained in the Work Plan.

Calibration will be documented in the Field Log Book as described in the SOPs attached to the Work Plan(refer to SOPs A7, A9, A10 and A14).

The Field Team Leader is responsible for maintaining the Field Meter Calibration Log for field instruments as part of field documentation in the Field Log Book (SOP A14). This log contains the same information as that for permanent lab instruments (ID number, name and model of

meter). The log also contains calibration results and notes for each day of equipment use. Documentation will include:

- Date of calibration
- Initials of person performing calibration
- Calibration results

Calibration will be performed daily prior to use for the multi-parameter probe and turbidity meter. In addition, the calibration will be checked any time the equipment is turned off during the day, any time there is a questionable response from the equipment and at the end of the day to determine if any drift has occurred. Changes in calibration will be noted on any boring logs or other sampling forms where field instrument readings were recorded that day. The thermometer will be calibrated prior to mobilization to the field.

9.2 LABORATORY INSTRUMENTS

Laboratory calibration procedures and frequency will be performed in accordance with the methods listed on Table 6-1. Documentation of calibration and maintenance is kept in bound notebooks. If equipment has been out of service for two weeks or longer, the equipment will be re-calibrated immediately prior to use and then at the frequencies prescribed below to ensure acceptable continual calibration while in use. Calibration procedures are summarized below (please refer to the LQMM in Appendix D for further information).

9.2.1 Volatile Organic Compounds (VOCs)

An instrument performance check solution must be performed at the beginning of each 12 hour period during which samples or standards are analyzed. The instrument performance checks, bromofluorobenzene for VOC analysis, must meet the criteria on Table 9-1.

A minimum of five concentrations of all compounds of interest is injected to determine response factors and define the linear range of the system. After an acceptable initial calibration has been performed, the calibration is verified at least once every 12 hours by analyzing a continuing calibration verification standard containing representative compounds from each fraction. The instrument is recalibrated (five standards) if the verification calibration check compounds are greater than $\pm 25\%$ of the initial calibration response.

9.2.2 <u>Metals</u>

For each analyte of interest, a calibration curve is prepared which covers the linear working range of the method. Two concentration levels and a blank are used to construct the calibration

curve, including one near the upper limit of the concentration range and one near the lower limit of the concentration range with an equitable distribution of the remaining points.

The linear regression technique is used to find a straight line that best fits a set of calibration points. For a calibration curve to be acceptable, the correlation coefficient must be greater than or equal to 0.995 and the difference between the value observed for each calibration standard and the value predicted for that standard using the fitted calibration curve must be within ten percent.

A calibration verification check sample and calibration blank are analyzed after every ten samples and at the end of the analyses for the day. Recalibration is performed if the ten percent limit is exceeded and the ten samples preceding the out-of-control check standard and blank are reanalyzed.

9.2.3 Cyanide

Total and WAD cyanides are based on the acid distillation of HCN from the sample using strong and weak acid respectively. The HCN is captured in a NaOH trap. The concentration of cyanide can be determined by a Titrimetric, Colorimetric, or potentiometric procedure. The procedure must address any interfering substances such as chlorine and sulfides. A calibration curve is run and this curve is verified by independent calibration check standards. If the standards are not within 15 % of the expected value, a new recalibration curve is required. Verify the calibration curve with every sample batch by analyzing a mid-range standard. Run one matrix spike sample for every 10 samples to check the efficiency of sample distillation. A matrix spike should be prepared by adding cyanide from the working standard or intermediate standard to 500 mL of sample to obtain a concentration of approximately 40 ug/L. Both the matrix and matrix spike duplicate are brought through the entire sample preparation and analytical process.

9.2.4 <u>Total Dissolved Solids and other Water Quality Parameters/Design Parameters</u>

The analytical balance must be calibrated at least annually by a certified calibration service. Auto-zero the balance and check level at least once daily, before first use. Calibration must be verified daily using a series of at least two National Institute for Standards and Technology (NIST) class "S" weights. Results must be within the instrument manufacturer's specifications. If results exceed the instrument specifications, the balance may not be used for analysis until it has been serviced and recalibrated. Please refer to the LQMM for further calibration information.

Water Quality Parameters/Design Parameters analyses vary in methodology and instrumentation, typically using ion chromatographs, spectrophotometers, analytical balances, etc. This leads to some variation in calibration for different analyses, but the general principles still apply. Calibration is always required, and consists of defining the working range by use of a

series of standard solutions. A minimum of five standards is typically used, and is verified on an ongoing basis (every ten to twenty samples at a minimum and at the end of the analysis sequence). Continuing calibrations are not performed for non-instrumental methods such as total dissolved solids (i.e., gravimetric methods), but balance weights are checked daily and documented in appropriate log books.

TABLE 9-1
INSTRUMENT PERFORMANCE CHECK CRITERIA FOR VOCs

| Bromofluorobenzene (BFB): | | |
|---------------------------|---|--|
| Mass | Intensity Required (relative abundance) | |
| 50 | 15 to 40% of mass 95 | |
| 75 | 30 to 60% of mass 95 | |
| 95 | base peak, 100% relative abundance | |
| 96 | 5 to 9% of mass 95 | |
| 173 | less than 2% of mass 174 | |
| 174 | greater than 50% of mass 95 | |
| 175 | 5 to 9% of mass 174 | |
| 176 | greater than 95% but less than 101% of mass 174 | |
| 177 | 5 to 9% of mass 176 | |

10. CORRECTIVE ACTION

A corrective action protocol that is both technically effective and administratively compatible is imperative to ensure accurate and timely correction of non-conformance with the Work Plan. To accomplish this, the organizational structure will be used (as described below) to initiate, implement and approve corrective actions.

10.1 FIELD PROCEDURES

The Field Team Leader is responsible for all site activities. In this role, the Field Team Leader at times is required to adjust the field program to accommodate site-specific needs. When it becomes necessary to modify the field program, the Field Team Leader will notify the PM (whom will notify the project team) of the anticipated change, document and implement the necessary changes after approval from the PM. Significant field changes may include: deleting a sampling location, moving a sample location, substituting different field equipment for that stated in the SSA or making any changes in the SSHP.

During the field investigation, each field team will be responsible for completing a DQCR. An example of a DQCR is provided in the appendices to this Work Plan. This report will include a description of any problems encountered and any corrective actions taken. These reports will be compiled and sent to the PM once every week if no problems or deviations from the schedule or Work Plan arise. However, should problems arise, the Field Team Leader will immediately notify the PM. DQCRs will be sent daily to the PM by fax or overnight delivery until the problem has been corrected.

Work may be stopped by the PM if corrective actions for significant problems are not effective and the problem remains unresolved.

10.2 LABORATORY PROCEDURES

Nonconformance is defined as any event which is beyond the limits established for laboratory performance, such as data which fall outside accepted bounds for accuracy and precision due to improper equipment calibration, maintenance operation or improper data verification. Any activity in the laboratory, which affects data quality, can result in a nonconformance.

Nonconformance associated with the statistical analysis and review of data is straightforward to identify. The Project Chemist will be responsible for the assessment of QC sample information. The PM will be notified of non-conformances that affect data quality to the degree that resampling may be necessary.

Corrective actions will be designed to correct the associated problems and to minimize the possibility of their recurrence. Examples of corrective actions include modifying nonconforming

procedures; tagging, repairing or replacing deficient equipment; training or replacing unqualified personnel; re-analyzing affected samples; marking rejected data; and, reissuing affected reports.

11. <u>DATA REDUCTION</u>

11.1 DATA REDUCTION

Upon completion of an analysis, the analyst will calculate the final sample results and associated QC results from the raw data. Additionally, for organic analyses, the analyst will review the scans of each sample and standard on the instrument terminal. The analyst will check for recovery of surrogate compounds, sample response within the linear range of the calibration curve and the integration of peaks in the chromatogram. Peaks that appear suspect may require re-integration by the analyst.

11.2 ANALYTICAL DATA PACKAGE REQUIREMENTS

The data will be submitted in both hard copy and on disk. For each analytical method run, the laboratory will report all analytes for each sample as a detected concentration or as less than the specific limits of quantitation. The data will be reported in units consistent with both Federal and State regulations, methods and guidelines. LRLs for all analytes will be recorded on all lab data report sheets for all samples within the sample set. Each analytical method run will be clearly identified as belonging to a specific analytical batch and will be reported with dates of collection, preparation and analysis; the laboratory also will report dilution factors for each sample. All soil samples will be reported on a dry-weight basis with percent moisture also reported. The laboratories will hold and make available all project raw data for a minimum period of ten years after samples have been analyzed.

A complete set of QC results, including calibration data, will be reported for each analytical batch. All required method QC procedures will be performed on project samples. A 45-calendar day turnaround time, from sample receipt of the last sample at the laboratory to data receipt by the Project Chemist, will be required for all samples.

11.2.1 <u>Laboratory Blank Samples</u>

All analytes will be reported for each laboratory blank. All sample results will be designated as corresponding to a particular laboratory blank in terms of analytical batch processing.

11.2.2 Surrogate Spike Samples

Surrogate spike recoveries will be reported with all organic method reports. Control limits for spike recoveries are identified on the Tables at the end of Section 6 and will be included with the analytical data package. The analytical data package also will specify the spiking concentration. Any out-of-control recoveries will result in the sample being re-prepared and re-analyzed as required by the analytical methods. Both sets of data will be reported, unless otherwise approved by the Project Chemist.

11.2.3 Matrix Spike/Matrix Spike Duplicate Samples

MS recoveries and RPDs will be reported for the samples. All sample results will be designated as corresponding to a particular MS/MSD pair. The report will indicate what field sample was spiked. MS/MSDs will be performed on project samples. The control limits for the MS/MSDs and RPD criteria are specified on the Tables at the end of Section 6 and will be included in the analytical data package.

11.2.4 Laboratory Control Samples/Laboratory Control Sample Duplicates

LCS results will be reported for the analyses. All sample results will be designated as corresponding to a particular LCS sample in terms of analytical batch processing. The control limits for the LCS/LCSDs and RPD criteria are specified on the Tables at the end of Section 6 and will be included in the analytical data package.

11.2.5 Other Method Quality Control

Additional QC checks, such as method of standard additions and serial dilutions for the analysis of metals, analytical spikes for metals, initial and continuing calibration data for all analytes, internal standard areas and tunes will be reported. Control limit ranges for all QC samples also will be reported in the analytical data package.

11.3 DATA PACKAGE ORGANIZATION

To aid data validation, separate data packages will be submitted for each type of analytical test as follows:

- VOCs:
- Metals;
- Cyanides;
- Water Quality Parameters.

Each data package will consist of the results for a maximum of 20 field samples. QC results discussed in Section 11.2 will be included for each batch. Laboratory Case Narratives, completed COC forms and cooler receipt forms will be submitted in the analytical data package.

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